

The Role of Exosomes in the Pathogenesis of Intrauterine Growth Restriction (IUGR): Future Perspective

Noushin Modaber¹, Roghayeh Amiri¹, Fatemeh Jayravand¹, Seyed Amir Hossein Hosseini², Bahareh Shateri Amiri³, Minoo Rostami⁴, Ezzat Khodashenas⁵, Bahareh Ghahrodizadehabyaneh⁶

¹ Shahid AkbarAbadi Clinical Research Development Unit (SHACRDU), School of Medicine, Iran University of Medical Sciences, Tehran, Iran.

² Department of Medical Genetics, Faculty of Medicine, Hormozgan University of Medical Sciences, Bandar Abbas, Iran.

³ Assistant Professor of Internal Medicine Department of Internal Medicine, School of Medicine Hazrat-e Rasool General Hospital Iran University of Medical Sciences, Mashhad, Iran.

⁴ Resident of Obstetrics and Gynecology, Medical Faculty, Shahid Beheshti University of Medical Sciences, Tehran, Iran.

⁵ Neonatal research center, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran.

⁶ Resident of Obstetrics and Gynecology, Medical Faculty, Tehran University of Medical Sciences, Tehran, Iran.

Abstract

Background: Exosomes are among the factors whose importance has been shown in many diseases today. Recently, it has been shown that exosomes play an important role in the pathogenesis of Intrauterine Growth Restriction (IUGR); however, few studies have been conducted in this regard.

Methods: The articles in this review study were retrieved from some databases including PubMed, Google scholar, and Scopus. All the included articles were in English, and those in other languages were excluded. Search keywords included IUGR, exosome, pathogenesis, Mechanism, Cell Signaling, Oxidative Stress, Inflammation, and Endothelial Dysfunction.

Results and conclusion: Studies have shown that exosomes contain factors, molecules and gene activators that affect molecular pathways regulation. These molecules play an important role in regulating inflammatory reactions, oxidative stress, and production of Reactive Oxygen Species (ROS). The activation of these pathways can aggravate the clinical symptoms of IUGR. In addition, exosomes can impress induction or inhibition of endothelial dysfunction, which leads to the development of IUGR. Hence, identifying upstream and downstream pathways helps design therapeutic strategies to treat patients.

Key Words: Exosome, Intrauterine Growth Restriction, Mechanism, Pathogenesis.

* Please cite this article as: Amiri R, Jayravand F, Modaber N, Hosseini SAH, Shateri Amiri B, Rostami M, Khodashenas E, Ghahrodizadehabyaneh B. The Role of Exosomes in the Pathogenesis of Intrauterine Growth Restriction (IUGR): Future Perspective. Int J Pediatr 2023; 11 (06):17979-17989. DOI: **10.22038/ijp.2023.72768.5284**

*Corresponding Author:

Bahareh Ghahrodizadehabyaneh, Resident of Obstetrics and Gynecology, Medical Faculty, Tehran University of Medical Sciences, Tehran, Iran. Email: school2022@gmail.com

Received date: Jun.01,2023; Accepted date:Jun.24,2023

1- INTRODUCTION

Intrauterine Growth Restriction (IUGR) is a condition in which the growth rate of the fetus decreases due to pathological factors (1, 2). The prevalence of this disease is different in different parts of the world, but on average, it happens in 5-20% of pregnancies (3). The clinical symptoms are organ failure, extreme thinness, paleness, and dry skin (4). Pathological factors that cause these conditions include maternal causes (blood pressure, diabetes, anemia, and malnutrition), fetal causes (genetic disorders such as aneuploidy, fetal infections, and congenital anomalies), and placental insufficiency (5).

On the other hand, exosomes are one of the causes of IUGR. Exosomes are a collection of extracellular vesicles with an average diameter of 100 nm. They originate from different organs and could transfer different molecules (6). Exosomes that originate from the placenta carry products, which affect various reproductive processes such as fetus and placenta development and also the adaptation of the mother and fetus during pregnancy; it is possible that if this system is affected, the IUGR phenomenon will occur (7, 8). In this study, we intend to investigate the role and function of exosomes in the pathogenesis of IUGR.

1-1. Search strategy

The articles in this review study were retrieved from some databases including PubMed, Google scholar, and Scopus. All the included articles were in English, and those in other languages were excluded. Search keywords included IUGR, exosome, pathogenesis, Mechanism, Cell Signaling, Oxidative Stress, Inflammation, and Endothelial Dysfunction.

1-2. Exosome and inflammation

Inflammation is one of the events that contributes to IUGR (9). CD81 increases

the expression of IL-6 by activating the JAK/STAT pathway. IL-6 increases the differentiation of naïve T cells to TH17, and suppresses Treg cells followed by increased inflammation. Increased expression of CD81 is seen in IUGR. Therefore, CD81 targeting can be considered as a therapeutic pathway (10). By inhibiting the JAK1/STAT3 pathway, CD63 reduces the expression of IL-6 and IL-27 followed by reduced inflammation. Decreased expression of CD63 has been observed in IUGR. Therefore, increasing the expression of CD63 can reduce inflammation and prevent the progression of IUGR (11).

The mir-1-3p suppresses the ETS1 gene by activating the TLR/MYD88 pathway. ETS1 prevents the interaction between NF-kB and CREB molecules; NF-kB and CREB binding increases the expression of MUC5AC, and subsequently promotes inflammation. On the other hand, by suppressing the ETS1 gene, mir1-3p facilitates the differentiation of naïve T cells into TH17 and finally causes inflammation. Increased expression of mir-1-3p is seen in IUGR. Therefore, targeting mir-1-3p can help in the treatment process of patients (12, 13). FASL, mir-520a, mir-16-5p, and mir-26-5p molecules reduce the NF-kB level by suppressing TLR/MYD88 pathway. FASL and mir-520a cause degradation of the P65 subunit of NF-kB and subsequently suppress inflammation (14, 15).

Mir-16-5P decreases PDCD4 expression by inhibiting the TLR/MYD88 pathway. PDCD4 activates the NF-kB and increases inflammation. Decreased expression of mir-16-5P has been reported in IUGR (16). Mir-26-5P reduces CTGF expression by inhibiting the TLR/MYD88 pathway. This gene is also one of the NF-kB activators (17). Mir-100-5P inhibits the mTOR pathway and reduces its expression. mTOR is one of the molecules that increases the process of protein synthesis

and energy production, and increasing the expression of this molecule causes the inflammation process to intensify. Therefore, reducing the expression of mTOR molecules, which occurs due to being targeted by mir-100-5P can be used as a therapeutic strategy for IUGR (18).

Mir-145-5p reduces the expression of TLR4 and MUC1 genes by inhibiting the TLR/MYD88 pathway. Decreased expression of Mir-145-5p has been observed in IUGR (19). Mir-125-5p, mir-146a-5P and mir-548e-5p molecules decrease TRAF6 gene expression by inhibiting the RAS/MAPK and TLR/MYD88 pathways. TRAF6 is one of the activators of NF- κ B and MAPK molecules. The expression of mir-125-5p, mir-146a, and mir-548e-5p decreases in IUGR; it can create a new way to treat IUGR by targeting the target molecules of these miRs (20, 21).

Mir-126-5p reduces HIF1 gene expression by inhibiting the PI3K/AKT pathway. On one hand, HIF1 activates NF- κ B and on the other hand, it increases the proliferation of immune system cells (22). Mir-195-5p reduces ATF6 gene expression by inhibiting the TLR/MYD88 pathway. ATF6 increases its expression in response to endoplasmic reticulum stress and subsequently increases TNF α expression and causes inflammation (23). Mir-199-5p reduces the expression by inhibiting the mTOR pathway. Rheb, directly, activates mTOR and increases the inflammation. Mir-199-5p expression is observed in IUGR (24).

Mir-210 decreases DR6 expression by inhibiting the TLR/MYD88 pathway. DR6 is an NF- κ B activator that increases inflammation (25, 26). Mir-221-3p reduces apoptosis and inflammation by inhibiting the CDKN1B (27). Mir-342-3p decreases AEG-1 expression by inhibiting the TLR/MYD88 pathway. AEG-1 is also a NF- κ B activator that can increase inflammation (28). Mir-518b decreases

RAP1B expression by inhibiting the RAS/MAPK pathway. RAP1B increases the migration of neutrophils and also the expression of IL-6. Decreased expression of mir-518b has been observed in IUGR (29, 30). In general, it can be concluded that inflammation has an important role in various diseases, including IUGR. Thus, we can take an important step in the diagnosis and treatment of IUGR by identifying molecules that can cause or increase inflammation.

1-3. Exosome and oxidative stress

One of the important factors in the IUGR occurrence is the increase of molecules resulting from the oxidative stress process during pregnancy. In addition to IUGR, ROS increment can affect the occurrence of other diseases such as preeclampsia and gestational diabetes. Therefore, we can identify the effective molecules and use them in the diagnosis, treatment, and prognosis determination (31). By inhibiting the RAS-MAPK pathway, mir-1-3p decreases IGF1 expression, and then increases ROS in the cells. Inhibiting mir-1-3p is one of the appropriate therapeutic tools, for example, a drug called propofol has been designed to inhibit this molecule and reduce the amount of ROS in cells to treat colorectal cancer (32, 33).

Mir-26-5p reduces ADAM17 gene expression by suppressing the notch-delta signaling pathway. This gene, under the influence of ROS molecules, causes further inflammation and cell damage. In this way, the observation of a decrease in the expression of mir-26-5p in IUGR can be understood and explained (34). Nox4 or NADPH OXIDASE 4 is one of the molecules increasing the amount of ROS in the cell under the influence of JAK-STAT and PI3K-AKT pathway. On the other hand, mir-100-5p by targeting the nox4 reduces ROS production. Mir-100-5p down expression can be seen both in the process of inflammation and production of oxidative substances (35).

Mir-195-5p, mir-145-5p, and mir-125-5p suppress VEGF and TGF β by inhibiting the PI3K pathway, which finally prevents the production of oxidant substances. TGF β can control ROS production directly or by reducing the antioxidant systems. Meanwhile, ROS elevation can also lead to TGF β up regulation. ROS elevation is also associated with VEGF increment; moreover, the VEGF molecule itself increases the amount of ROS in the cells by inducing the angiogenesis process (36, 37).

One of the important mechanisms of the cell to deal with oxidative substances is to activate the NRF2/SIRT1 signaling pathway. The activation of NRF2 increases the production of effective proteins on oxidant inhibition. Mir-126-3p reduces oxidants by activating the SIRT1/Nrf2 signaling pathway. Decreased expression of this molecule is observed in IUGR (38). Mir-199-5p decreases CAV1 expression through activating the NRF2 pathway. The CAV1 induces cell autophagy under the oxidative stress conditions, while mir-199-5p activates NRF2 by inhibiting the CAV1; as mentioned in the previous section, NRF2 activation increases the production of antioxidant proteins.

One of the weapons of war and bioterrorism is chemical substance (Sulfur mustard). This molecule can cause fatal damage to the body by increasing the oxidants in the lungs. Increasing the expression of mir-199-5p can significantly reduce this mortal damage, which proves the therapeutic role of this molecule (39). Mir-103-3p and mir-210 decrease the expression of BNIP3 through inhibiting the autophagy pathway. In line with ROS elevation, BNIP3 is activated as a sensor and moves the cell towards autophagy. Therefore, ROS increment is associated with BNIP3 up-regulation and mir-103-3p and mir-210 downregulation. Mir-210, mir-103-3p, and BNIP3 can be used as determining factors in detecting the

progress in the oxidative stress process. Decreased expression of mir-210 and mir-103-3p has been observed in IUGR (40-42).

Mir-221-3p down regulates the HIF1 α expression through inhibiting the PI3K/AKT pathway. The HIF1 α gene can cause angiogenesis by activating the VEGF, and inflammation in different tissues by activating the NF- κ B. In this way, increasing the expression of mir-221-3p can play a therapeutic role in IUGR (43). Mir-517a downregulates CDKN1C expression through inhibiting the JNK pathway. Silencing the mir-517a molecule increases the expression of CDKN1C, increases cell proliferation, and consequently increases the amount of ROS in the cells. The expression of HIF1 α increases under hypoxia conditions (44). On the other hand, it can act as a transcription factor for many other molecules, including HLA-G. Researchers have observed a link between hydrogen peroxide increment and HLA-G; so, it can be concluded that the increased level of HLA-G can be used as a diagnostic or therapeutic strategy (45, 46).

1-4. LncRNAs and endothelial dysfunction

One of the important causes of IUGR is endothelial cell (EC) dysfunction. It is also effective in preeclampsia and other complications caused by placental syndrome. Therefore, it is important to identify the molecular pathways involved in EC dysfunction in the process of timely diagnosis, prevention, or treatment of the disease (47).

The exosome derived from platelet-rich plasma (PRP-exosome) leads to dysfunction of retinal vascular endothelium through activation of TLR4 signaling pathway. TLR4 signaling in EC may be stimulated by LPS and oxidized phospholipids and can lead to endothelial activation and inflammation. LPS leads to

the activation of TLR4 through the signaling pathways of NADPH oxidase, ROS, Endothelial nitric oxide synthase (eNOS), MAPK, and NF κ B, which lead to inflammation and EC dysfunction. This mechanism is also effective in placental vascular endothelium disorder and IUGR (48-50).

Increased expression of miR-143/145 by activating the TGF β 1 signaling pathway causes disruption in the differentiation and function of pulmonary artery smooth muscle cells (PASMC); it is also a possible mechanism for increasing the pulmonary artery pressure in patients. TGF β 1 exerts strong effects on EC and SMCs. The release of large amounts of TSP1 (thrombospondin 1) and TGF β 1 from activated platelets through an increased response to collagen may lead to increased proliferation of EC. Inhibitors of TSP1 or its receptor CD47 may stop aberrant cell proliferation in the vascular endothelium. In general, the role of TSP1 and galectin-3 in inducing EC dysfunction through TGF β 1 signaling pathway can be used in the treatment of patients. TGF β 1 reduces the production of NOS and instead increases the production of endothelial NOS. Increased expression of mir-143 is seen in the vascular smooth muscle layer of the patients with increased pulmonary artery pressure. The same mechanism can lead to placental vascular EC dysfunction and cause IUGR (51, 52).

Trimethylamine-N-oxidase (TMAO) is a new factor causing inflammation and EC dysfunction. By stimulating the cell, it triggers exosome production; it also leads to the induction of cell inflammation and apoptosis through NF- κ B signaling pathway activation. This signaling pathway is activated by the advanced glycation end product (AGE) receptor and is a key regulator of inflammation and oxidative stress. By stimulating the oxidative stress, it leads to the dysfunction of placental vascular EC and lays the

foundation of IUGR creation. On the other hand, inhibiting this signaling pathway limits the vicious cycle of inflammation and oxidative stress; this mechanism can be used for therapeutic purposes.

Various studies have shown the impressive role of AGE receptors in cellular processes, such as inflammation, degradation, and oxidative stress (53-55). The death of ECs leads to the release of apoptotic-exosome like vesicles (APOExo). APOExo regulates the survival of EC and affects migration, angiogenesis, and cell differentiation. ECs that are exposed to APOExo through the activation of NF- κ B signaling pathway show reduced levels of apoptosis and angiogenesis activity; it also leads to dedifferentiation and gradual loss of ECs related markers as well as acquirement of Mesenchymal cell (S100A4, α SMA) markers. Finally, vascular dysfunction occurs. NF- κ B silencing reverses the anti-apoptotic and pro-migratory effects and prevents the angiostatic properties and CD31 downregulation in ECs exposed to ApoExo. ApoExo has been identified as a novel inducer of NF- κ B activation in ECs; it shows the pivotal role of this signaling pathway in coordination with the ApoExo-induced functional changes in ECs. Hence, targeting the ApoExo-mediated NF- κ B activation in ECs provides novel avenues to prevent EC dysfunction (56). Activation of this signaling pathway in pregnant women leads to IUGR and other disorders related to placental vessels (56, 57).

miR-34a leads to dysfunction of vascular endothelium and acceleration of end organ damage through the oxidant-sensitive mechanism and reducing the activity of Sirt 1 signaling pathway. Activation of SIRT1 by 1720SRT ameliorates EC dysfunction with aging through increasing the COX-2 signaling and reducing the oxidative stress and inflammation. Specific activation of SIRT1 is a promising therapeutic strategy for age-related EC

dysfunction in humans. Oxidative stress is the main mechanism of this signaling pathway in placental vascular endothelial dysfunction (58).

Microtubulin inhibitor MT189 suppresses angiogenesis by reducing the EC proliferation, migration, and differentiation through the JNK-VEGF/VEGFR-2 signaling axis. miR-210 also leads to EC protection against oxidative stress through the activation of the VEGF/VEGFR2 signaling pathway. Neural Progenitor Cells (NPCs) protect ECs against oxidative stress caused by AngII, which leads to apoptosis and EC dysfunction. As a therapeutic target, this mechanism can be used in the IUGR treatment and reduce the effect of oxidative stress caused by AngII in the placental vascular endothelium (59).

Exosomal miR424-5p induced by the endoplasmic reticulum-stressed head and neck squamous cell carcinoma leads to the inhibition of angiogenesis and migration of ECs through the inactivation of the Wnt/B catenin signaling pathway. IL-8, a known

angiogenesis factor, is also a transcriptional target of the Wnt/B catenin signaling pathway in ECs. Expression of Wnt-1 or B-cateninS37A induces IL-8 expression. Therefore, it could be concluded that the cited signaling pathway probably induces angiogenesis through the induction of known angiogenesis regulators such as IL-8. A similar mechanism can play a role in placental EC dysfunction and IUGR (**Table 1**) (60, 61). IL-1-associated Kinase-1 (IRAK-1) and TNF receptor-associated factor 6 (TRAF6) have been identified as the direct targets of mir-155. It can directly bind to the NF-kB P65.

All these target genes are important mediators of inflammation and EC dysfunction. In fact, Exosomal mir-155 also leads to endothelial and mitochondrial cell dysfunction by activating the NF-kB signaling pathway; it could be also useful as a therapeutic target for pregnancies complicated by IUGR and other disorders related to placental vessels (62).

Table-1: Potential therapeutic mechanisms of exosome from different sources

Source	Mechanism	Ref.
umbilical cord blood	Increased expression of mir-150 causing angiogenesis	(63)
pulmonary vascular endothelial cells and pulmonary arterial smooth muscle cells	mir-214-3p, mir-326-3p, and mir-125b-2-3p inhibition FoxM1 expression and decreased pulmonary hypertension in IUGU	(64)
fibroblast	mir-200 expression causing angiogenesis	(65, 66)
Mesenchymal cell	mir-20 expression causing angiogenesis	(67, 68)

2- CONCLUSION

In general, exosomes contain many molecules, factors, and regulators of gene expression, each of which regulates different molecular pathways. However, most of them are involved in inflammation induction and ROS production as well as endothelial dysfunction. These factors can contribute to the exacerbation of IUGR.

Hence, the identification of upstream and downstream pathways can play an important role in designing therapeutic strategies.

3- REFERENCES

1. Romo A, Carceller R, Tobajas J. Intrauterine growth retardation (IUGR): epidemiology and etiology. *Pediatr Endocrinol Rev.* 2009; 6 Suppl 3:332-6.

2. Vandenbosche RC, Kirchner JT. Intrauterine growth retardation. *Am Fam Physician*. 1998; 58(6):1384-90, 93-4.
3. Saleem T, Sajjad N, Fatima S, Habib N, Ali SR, Qadir M. Intrauterine growth retardation - small events, big consequences. *Italian Journal of Pediatrics*. 2011; 37(1):41.
4. Kesavan K, Devaskar SU. Intrauterine Growth Restriction: Postnatal Monitoring and Outcomes. *Pediatr Clin North Am*. 2019; 66(2):403-23.
5. Ergaz Z, Avgil M, Ornoy A. Intrauterine growth restriction-etiology and consequences: what do we know about the human situation and experimental animal models? *Reprod Toxicol*. 2005; 20(3):301-22.
6. Bai K, Li X, Zhong J, Ng EHY, Yeung WSB, Lee CL, Chiu PCN. Placenta-Derived Exosomes as a Modulator in Maternal Immune Tolerance During Pregnancy. *Front Immunol*. 2021; 12:671093.
7. Miranda J, Paules C, Nair S, Lai A, Palma C, Scholz-Romero K, Rice GE, Gratacos E, Crispi F, Salomon C. Placental exosomes profile in maternal and fetal circulation in intrauterine growth restriction - Liquid biopsies to monitoring fetal growth. *Placenta*. 2018; 64:34-43.
8. Hakemi MS, Nassiri AA, Nobakht A, Mardani M, Darazam IA, Parsa M, Miri MM, Shahrami R, Ahmadi Koomleh A, Entezarmahdi K, Karimi A. Benefit of hemoadsorption therapy in patients suffering sepsis-associated acute kidney injury: a case series. *Blood Purification*. 2022; 51(10):823-30.
9. Shalom-Paz E, Weill S, Ginzberg Y, Khatib N, Anabusi S, Klorin G, Sabo E, Beloosesky R. IUGR induced by maternal chronic inflammation: long-term effect on offspring's ovaries in rat model-a preliminary report. *J Endocrinol Invest*. 2017; 40(10):1125-31.
10. Ding H, Dai Y, Lei Y, Wang Z, Liu D, Li R, Shen L, Gu N, Zheng M, Zhu X, Zhao G, Hu Y. Upregulation of CD81 in trophoblasts induces an imbalance of Treg/Th17 cells by promoting IL-6 expression in preeclampsia. *Cell Mol Immunol*. 2019; 16(1):302-12.
11. Li Y, Yan C, Fan J, Hou Z, Han Y. MiR-221-3p targets Hif-1 α to inhibit angiogenesis in heart failure. *Laboratory Investigation*. 2021; 101(1):104-15.
12. Li L, Ma X, Zhao YF, Zhang C. MiR-1-3p facilitates Th17 differentiation associating with multiple sclerosis via targeting ETS1. *Eur Rev Med Pharmacol Sci*. 2020; 24(12):6881-92.
13. Song KS, Yoon JH, Kim KS, Ahn DW. c-Ets1 inhibits the interaction of NF- κ B and CREB, and downregulates IL-1 β -induced MUC5AC overproduction during airway inflammation. *Mucosal Immunol*. 2012; 5(2):207-15.
14. Ariyakumar G, Morris JM, McKelvey KJ, Ashton AW, McCracken SA. NF- κ B regulation in maternal immunity during normal and IUGR pregnancies. *Sci Rep*. 2021; 11(1):20971.
15. Wang R, Zhao Z, Zheng L, Xing X, Ba W, Zhang J, Huang M, Zhu W, Liu B, Meng X, Bai J, Li C, Li H. MicroRNA-520a suppresses the proliferation and mitosis of HaCaT cells by inactivating protein kinase B. *Exp Ther Med*. 2017;14(6):6207-12.
16. Liang X, Xu Z, Yuan M, Zhang Y, Zhao B, Wang J, Zhang A, Li G. MicroRNA-16 suppresses the activation of inflammatory macrophages in atherosclerosis by targeting PDCD4. *Int J Mol Med*. 2016; 37(4):967-75.
17. Sánchez-López E, Rayego S, Rodrigues-Díez R, Rodríguez JS, Rodrigues-Díez R, Rodríguez-Vita J, Carvajal G, Aroeira LS, Selgas R, Mezzano SA, Ortiz A, Egido J, Ruiz-Ortega M. CTGF promotes inflammatory

cell infiltration of the renal interstitium by activating NF-kappaB. *J Am Soc Nephrol*. 2009; 20(7):1513-26.

18. Luo P, Jiang C, Ji P, Wang M, Xu J. Exosomes of stem cells from human exfoliated deciduous teeth as an anti-inflammatory agent in temporomandibular joint chondrocytes via miR-100-5p/mTOR. *Stem Cell Res Ther*. 2019; 10(1):216.

19. Jara D, Carvajal P, Castro I, Barrera MJ, Aguilera S, González S, Molina C, Hermoso M, González MJ. Type I Interferon Dependent hsa-miR-145-5p Downregulation Modulates MUC1 and TLR4 Overexpression in Salivary Glands From Sjögren's Syndrome Patients. *Front Immunol*. 2021; 12:685837.

20. Paik JH, Jang JY, Jeon YK, Kim WY, Kim TM, Heo DS, Kim TM, Heo DS, Kim CW. MicroRNA-146a downregulates NFκB activity via targeting TRAF6 and functions as a tumor suppressor having strong prognostic implications in NK/T cell lymphoma. *Clin Cancer Res*. 2011; 17(14):4761-71.

21. Rasheed Z, Rasheed N, Abdulmonem WA, Khan MI. Author Correction: MicroRNA-125b-5p regulates IL-1β induced inflammatory genes via targeting TRAF6-mediated MAPKs and NF-κB signaling in human osteoarthritic chondrocytes. *Sci Rep*. 2019; 9(1):14729.

22. Liu J, Wei E, Wei J, Zhou W, Webster KA, Zhang B, Li D, Zhang G, Wei Y, Long Y, Qi X, Zhang Q, Xu D. MiR-126-HMGB1-HIF-1 Axis Regulates Endothelial Cell Inflammation during Exposure to Hypoxia-Acidosis. *Dis Markers*. 2021; 2021:4933194.

23. Scalavino V, Piccinno E, Bianco G, Schena N, Armentano R, Giannelli G, Serino G. The Increase of miR-195-5p Reduces Intestinal Permeability in Ulcerative Colitis, Modulating Tight Junctions' Expression. *Int J Mol Sci*. 2022; 23(10).

24. Wang Y, Luo J, Wang X, Yang B, Cui L. MicroRNA-199a-5p Induced Autophagy and Inhibits the Pathogenesis of Ankylosing Spondylitis by Modulating the mTOR Signaling via Directly Targeting Ras Homolog Enriched in Brain (Rheb). *Cell Physiol Biochem*. 2017; 42(6):2481-91.

25. Zaccagnini G, Greco S, Longo M, Maimone B, Voellenkle C, Fuschi P, Carrara M, Creo P, Maselli D, Tirone M, Mazzone M, Gaetano C, Spinetti G, Martelli F. Hypoxia-induced miR-210 modulates the inflammatory response and fibrosis upon acute ischemia. *Cell Death Dis*. 2021; 12(5):435.

26. Zhang D, Cao X, Li J, Zhao G. MiR-210 inhibits NF-κB signaling pathway by targeting DR6 in osteoarthritis. *Sci Rep*. 2015; 5:12775.

27. Yang H, Zhang L, Wang Q. MicroRNA-221-3p alleviates cell apoptosis and inflammatory response by targeting cyclin dependent kinase inhibitor 1B in chronic obstructive pulmonary disease. *Bioengineered*. 2021; 12(1):5705-15.

28. Zhang S, Liu L, Lv Z, Li Q, Gong W, Wu H. MicroRNA-342-3p Inhibits the Proliferation, Migration, and Invasion of Osteosarcoma Cells by Targeting Astrocyte-Elevated Gene-1 (AEG-1). *Oncol Res*. 2017; 25(9):1505-15.

29. Konwar C, Manokhina I, Terry J, Inkster AM, Robinson WP. Altered levels of placental miR-338-3p and miR-518b are associated with acute chorioamnionitis and IL6 genotype. *Placenta*. 2019; 82:42-5.

30. Eshraghi N, Jamal A, Eshraghi N, Kashanian M, Sheikhsari N. Cerebroplacental ratio (CPR) and reduced fetal movement: predicting neonatal outcomes. *The Journal of Maternal-Fetal & Neonatal Medicine*. 2022; 35(10):1923-8.

31. Rudov A, Balduini W, Carloni S, Perrone S, Buonocore G, Albertini MC. Involvement of miRNAs in placental alterations mediated by oxidative stress. *Oxid Med Cell Longev*. 2014; 2014:103068.
32. Ye LL, Cheng ZG, Cheng XE, Huang YL. Propofol regulates miR-1-3p/IGF1 axis to inhibit the proliferation and accelerates apoptosis of colorectal cancer cells. *Toxicol Res (Camb)*. 2021; 10(4):696-705.
33. Kashanian M, Eshraghi N, Sheikhsari N, Eshraghi N. Comparing the efficacy of dilapan with extra-amniotic saline infusion and oral misoprostol for cervical ripening in term pregnancies. *The Journal of Maternal-Fetal & Neonatal Medicine*. 2022; 35(25):5616-20.
34. Wen X, Yin Y, Li X, He T, Wang P, Song M, Gao J. Effect of miR-26a-5p targeting ADAM17 gene on apoptosis, inflammatory factors and oxidative stress response of myocardial cells in hypoxic model. *J Bioenerg Biomembr*. 2020; 52(2):83-92.
35. Li X, Wang Y, Cai Z, Zhou Q, Li L, Fu P. Exosomes from human umbilical cord mesenchymal stem cells inhibit ROS production and cell apoptosis in human articular chondrocytes via the miR-100-5p/NOX4 axis. *Cell Biol Int*. 2021; 45(10):2096-106.
36. Barutta F, Bellini S, Guarrera S, Matullo G, Schalkwijk C, Stehouwer CD, Chaturvedi N, Soedamah-Muthu SS, Durazzo M, Gruden G. Association of serum MicroRNA-145-5p levels with microvascular complications of type 1 Diabetes: The EURODIAB prospective complications study. *Diabetes Res Clin Pract*. 2022; 190:109987.
37. Ye Y, Liu Q, Li C, He P. miR-125a-5p Regulates Osteogenic Differentiation of Human Adipose-Derived Mesenchymal Stem Cells under Oxidative Stress. *Biomed Res Int*. 2021; 2021:6684709.
38. Li J, Yang C, Wang Y. miR-126 overexpression attenuates oxygen-glucose deprivation/reperfusion injury by inhibiting oxidative stress and inflammatory response via the activation of SIRT1/Nrf2 signaling pathway in human umbilical vein endothelial cells. *Mol Med Rep*. 2021; 23(2).
39. Gong C, Xu Q, Zhang X, Mao G, Pei Z, Meng W, Cen JF, He XW, Lu Y, Xu QQ, Xiao K. HMSCs exosome-derived miR-199a-5p attenuates sulfur mustard-associated oxidative stress via the CAV1/NRF2 signaling pathway. *Research Square*; 2022.
40. Diao H, Liu B, Shi Y, Song C, Guo Z, Liu N, Song X, Lu Y, Lin X, Li Z. MicroRNA-210 alleviates oxidative stress-associated cardiomyocyte apoptosis by regulating BNIP3. *Biosci Biotechnol Biochem*. 2017; 81(9):1712-20.
41. Wang Y, Song X, Li Z, Liu N, Yan Y, Li T, Li T, Sun W, Guan Y, Li M, Yang Y, Yang X, Liu B. MicroRNA-103 Protects Coronary Artery Endothelial Cells against H₂O₂-Induced Oxidative Stress via BNIP3-Mediated End-Stage Autophagy and Antipyroptosis Pathways. *Oxid Med Cell Longev*. 2020; 2020:8351342.
42. Sheikh M, Ostadrahimi P, Salarzaei M, Parooie F. Cardiac complications in pregnancy: a systematic review and meta-analysis of diagnostic accuracy of BNP and N-terminal pro-BNP. *Cardiology and Therapy*. 2021; 10:501-14.
43. Fu M, Zhu Y, Zhang J, Wu W, Sun Y, Zhang X, Tao J, Li Z. MicroRNA-221-3p Suppresses the Microglia Activation and Seizures by Inhibiting of HIF-1 α in Valproic Acid-Resistant Epilepsy. *Frontiers in Pharmacology*. 2021; 12.
44. Yang C, Yan Z, Hu F, Wei W, Sun Z, Xu W. Silencing of microRNA-517a induces oxidative stress injury in

melanoma cells via inactivation of the JNK signaling pathway by upregulating CDKN1C. *Cancer Cell International*. 2020; 20(1):32.

45. Garziera M, Scarabel L, Toffoli G. Hypoxic Modulation of HLA-G Expression through the Metabolic Sensor HIF-1 in Human Cancer Cells. *J Immunol Res*. 2017; 2017:4587520.

46. Zhou X, Zhang GY, Wang J, Lu SL, Cao J, Sun LZ. A novel bridge between oxidative stress and immunity: the interaction between hydrogen peroxide and human leukocyte antigen G in placental trophoblasts during preeclampsia. *Am J Obstet Gynecol*. 2012; 206(5):447.e7-16.

47. Yinon Y, Kingdom JC, Odutayo A, Moineddin R, Drewlo S, Lai V, Lai V, Cherney DZI, Hladunewich MA. Vascular dysfunction in women with a history of preeclampsia and intrauterine growth restriction: insights into future vascular risk. *Circulation*. 2010; 122(18):1846-53.

48. Zhang W, Dong X, Wang T, Kong Y. Exosomes derived from platelet-rich plasma mediate hyperglycemia-induced retinal endothelial injury via targeting the TLR4 signaling pathway. *Experimental eye research*. 2019; 189:107813.

49. Sheikh M, Mahabadi BS, Ostadrahimi P. Infective endocarditis in Iranian children: A systematic review and meta-analysis in three age groups. *Int J Adv Res Biol Sci*. 2019; 6(5):110-7.

50. Zadeh FJ, Ghasemi Y, Bagheri S, Maleknia M, Davari N, Rezaeeyan H. Do exosomes play a role in cardiovascular disease development in hematological malignancy? *Molecular Biology Reports*. 2020; 47:5487-93.

51. Deng L, Blanco FJ, Stevens H, Lu R, Caudrillier A, McBride M, McClure JD, Grant J, Thomas M, Frid M, Stenmark K, White K, Seto AG, Morrell NW, Bradshaw AC, MacLean MR, Baker AH. MicroRNA-143 activation regulates

smooth muscle and endothelial cell crosstalk in pulmonary arterial hypertension. *Circulation research*. 2015; 117(10):870-83.

52. Mehrpisheh S, Mosayebi Z, Memarian A, Kadivar M, Nariman S, Ostadrahimi P, Dalili H. Evaluation of specificity and sensitivity of gastric aspirate shake test to predict surfactant deficiency in Iranian premature infants. *Pregnancy Hypertension: An International Journal of Women's Cardiovascular Health*. 2015; 5(2):182-6.

53. Liu X, Shao Y, Tu J, Sun J, Li L, Tao J, Chen J. Trimethylamine-n-oxide-stimulated hepatocyte-derived exosomes promote inflammation and endothelial dysfunction through nuclear factor-kappa b signaling. *Annals of Translational Medicine*. 2021; 9(22).

54. Xiong Y, Chen L, Yan C, Zhou W, Endo Y, Liu J, Hu L, Hu Y, Mi B, Liu G. Circulating exosomal miR-20b-5p inhibition restores Wnt9b signaling and reverses diabetes-associated impaired wound healing. *Small*. 2020; 16(3):1904044.

55. Shahramian I, Tabrizian K, Ostadrahimi P, Afshari M, Soleymanifar M, Bazi A. Therapeutic effects of ursodeoxycholic acid in neonatal indirect hyperbilirubinemia: a randomized double-blind clinical trial. *Archives of Anesthesiology and Critical Care*. 2019; 5(3):99-103.

56. Migneault F, Dieudé M, Turgeon J, Beillevaire D, Hardy M-P, Brodeur A, Thibodeau N, Perreault C, Hébert MJ. Apoptotic exosome-like vesicles regulate endothelial gene expression, inflammatory signaling, and function through the NF- κ B signaling pathway. *Scientific reports*. 2020; 10(1):1-15.

57. Zadeh FJ, Akbari T, Zayeri ZD, Samimi A, Davari N, Rezaeeyan H. The role of molecular mechanism of Ten-

- Eleven Translocation2 (TET2) family proteins in pathogenesis of cardiovascular diseases (CVDs). *Molecular Biology Reports*. 2020; 47(7):5503-9.
58. Ibrahim AA, Wahby AA, Ashmawy I, Saleh RM, Soliman H. Association of exosomal miR-34a with markers of dyslipidemia and endothelial dysfunction in children and adolescents with T1DM. *Journal of Clinical Research in Pediatric Endocrinology*. 2020; 12(4):401.
59. Liu H, Wang J, Chen Y, Chen Y, Ma X, Bihl JC, Yang Y. NPC-EXs alleviate endothelial oxidative stress and dysfunction through the miR-210 downstream Nox2 and VEGFR2 pathways. *Oxidative Medicine and Cellular Longevity*. 2017; 2017.
60. Wang Z, Jiao P, Zhong Y, Ji H, Zhang Y, Song H, Du H, Ding X, Wu H. The Endoplasmic Reticulum-Stressed Head and Neck Squamous Cell Carcinoma Cells Induced Exosomal miR-424-5p Inhibits Angiogenesis and Migration of Humanumbilical Vein Endothelial Cells Through LAMC1-Mediated Wnt/ β -Catenin Signaling Pathway. *Cell Transplantation*. 2022; 31:09636897221083549.
61. Akhavan S, Tutunchi S, Malmir A, Ajorlou P, Jalili A, Panahi G. Molecular study of the proliferation process of beta cells derived from pluripotent stem cells. *Molecular Biology Reports*. 2022:1-8.
62. Ge X, Tang P, Rong Y, Jiang D, Lu X, Ji C, Wang J, Huang C, Duan A, Liu Y, Chen X, Chen X, Xu Z, Wang F, Wang Z, Li X, Zhao W, Fan J, Liu W, Yin G, Cai W. Exosomal miR-155 from M1-polarized macrophages promotes EndoMT and impairs mitochondrial function via activating NF- κ B signaling pathway in vascular endothelial cells after traumatic spinal cord injury. *Redox biology*. 2021; 41:101932.
63. Luo J, Fan Y, Shen L, Niu L, Zhao Y, Jiang D, Zhu L, Jiang A, Tang Q, Ma J, Jin L, Wang J, Li X, Zhang S, Zhu L. The Pro-angiogenesis Of Exosomes Derived from Umbilical Cord Blood of Intrauterine Growth Restriction Pigs Was Repressed Associated with MiRNAs. *Int J Biol Sci*. 2018; 14(11):1426-36.
64. Luo X, Hang C, Zhang Z, Le K, Ying Y, Lv Y, Lv Y, Yan L, Huang Y, Ye L, Xu X, Zhong Y, Du L. PVECs-Derived Exosomal microRNAs Regulate PSMCs via FoxM1 Signaling in IUGR-induced Pulmonary Hypertension. *Journal of the American Heart Association*. 2022; 11(24):e027177.
65. Hu T-X, Wang G, Guo X-J, Sun Q-Q, He P, Gu H, Huang Y, Gao L, Ni X. MiR 20a,-20b and -200c are involved in hydrogen sulfide stimulation of VEGF production in human placental trophoblasts. *Placenta*. 2016; 39:101-10.
66. Ranjan P, Kumari R, Goswami SK, Li J, Pal H, Suleiman Z, Cheng Z, Krishnamurthy P, Kishore R, Verma SK. Myofibroblast-derived exosome induces cardiac endothelial cell dysfunction. *Frontiers in Cardiovascular Medicine*. 2021; 8:676267.
67. Hu T-X, Wang G, Guo X-J, Sun Q-Q, He P, Gu H, Huang Y, Gao L, Ni X. MiR 20a,-20b and -200c are involved in hydrogen sulfide stimulation of VEGF production in human placental trophoblasts. *Placenta*. 2016; 39:101-10.
68. Zhang L, Song Y, Chen L, Li D, Feng H, Lu Z, Fan T, Chen Z, Livingston MJ, Geng Q. MiR-20a-containing exosomes from umbilical cord mesenchymal stem cells alleviates liver ischemia/reperfusion injury. *Journal of Cellular Physiology*. 2020; 235(4):3698-710.