

Is Low Intensity Pulsed Ultrasound (LIPUS) Effective in the Success Rate of Alveolar Cleft Graft?

Arezoo Jahanbin¹, * Fahimeh Farzanegan², Amirreza Mashreghi³, Seyed Mohammad Ali Raisolsadat⁴, Sayed Hosein Hoseini Zarch⁵, Neda Mostafaee⁶

¹ Professor of Department of Orthodontics, School of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran.

² Associate Professor of Department of Orthodontics, School of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran.

³ Assistant professor of Department of Orthodontics, School of Dentistry, Zahedan University of Medical Sciences, Zahedan, Iran.

⁴ Assistant professor of Department of Pediatric Surgery, School of Medicine, Azad University of Mashhad, Mashhad, Iran.

⁵ Associate Professor of the Department of Oral and Maxillofacial Radiology, School of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran.

⁶ Assistant professor of Department of Physical Therapy , School of Paramedical Sciences, Mashhad University of Medical Sciences , Mashhad, Iran.

Abstract

Background: The aim of this study was to investigate the efficacy of the low intensity pulsed ultrasound in reconstruction of the alveolar cleft area after autologous bone grafting.

Methods: In this study, 14 patients with unilateral or bilateral cleft lip and palate aged between 9 to 13 years, were selected. Seven of the patients received only the autologous bone graft and the remaining seven underwent alveolar bone graft, and one week after transplantation were subjected to LIPUS waves for five minutes at a frequency of 1 MHz and 100 mW in the area of the graft for a period of five weeks (15 sessions). CBCT images were immediately taken after surgery and three months later. In CBCTs, bone mass was measured with two components of height and bone thickness and the quality was measured by evaluating the bone density by means of the Hounsfield Uniform (HU) mean. Data analysis was done via SPSS version 16 software and using paired t, independent t, and Mann-Whitney and Wilcoxon tests. A significance level of 0.05 was considered.

Results: The mean changes of the sagittal thickness (P=.944), sagittal height (P=.482), and axial thickness (P=.242) before and after surgery, in contrast to the axial height (P=.357) and density (P=.443), were less in the control group than the intervention group, but the differences were not significant for any of variables. In the intervention and control groups, in comparison to the immediate results after surgery, the mean values of the sagittal thickness, sagittal height, axial thickness, and axial height decreased significantly three months later; but the mean loss in density was not significant.

Conclusion: Ultrasound in repairing alveolar defect in patients with cleft palate has no significant effect on clinical success criteria.

Key Words: Alveolar bone graft, Cleft lip and palate, Low intensity pulsed ultrasound.

<u>* Please cite this article as</u>: Jahanbin A, Farzanegan F, Mashreghi A, Raisolsadat SMA, Hoseini Zarch SH, Mostafaee N. Is Low Intensity Pulsed Ultrasound (LIPUS) Effective in the Success Rate of Alveolar Cleft Graft? Int J Pediatr 2022; 10 (12):17093-17102. DOI: **10.22038/ijp. 2022.66844.5008**

Received date: jul.20,2022; Accepted date:Oct.20,2022

^{*}Corresponding Author:

Fahimeh Farzanegan, Associate Professor of Department of Orthodontics, School of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran. Email: farzaneganf@mums.ac.ir

1- INTRODUCTION

Orofacial clefts are one of the most common abnormalities in the craniofacial area (1). In general, the incidence of cleft lip and palate in different societies vary from 1 to 1.5 in 1,000 births (2, 3). Bone graft in the gap region is used as a common technique for repairing defects in patients with cleft lips and palates; however, there is uncertainty about the right time to do this (4). Sometime initial bone grafting is performed at the same time as lip reconstruction before the age of two years, but satisfactory results from this technique do not exist at this age range (5, 6). In the mixed dentition procedure, the secondary bone graft is performed within the age range of 7 to 12 years (7-14). In different studies, the use of alloplastic (12) and autogenous (6-20) transplants for the reconstruction of alveolar clefts has been studied and compared (12).

Among the methods used to better reconstruct bone tissue, low intensity pulsed ultrasound (LIPUS) has been widely used in medicine as a therapeutic tool (21). Unlike medical imaging (which transmits ultrasound waves to tissue and processes a reversible waveform for image production), ultrasound therapy is a oneway energy application that is performed by an appliance of an audio device with a frequency of 1 to 3 MHz and an intensity between 0.1 to 3 watts per square centimeter (22). The ultrasound energy causes the molecules to melt through acoustic waves. This increased molecular movement causes frictional heat and, as a result, an increase in the temperature of the tissue. These effects of ultrasound are said to increase the collagen's flexibility, increase the speed of nervous conduction, change in local perfusion, increase enzyme activity, change muscle contractility activity, and increase the nociceptive threshold (23). Ultrasound is often used for the benefits of heat generation, while some recent researches in this field have

indicated that the non-thermal effects of ultrasound are also effective and even predominant (24). The mechanisms that are thought to be effective in generating non-thermal effects these include cavitation and acoustic streaming or micro-streaming. It has been said that phenomena these increase cell permeability and influence the process of cellular growth and thus improve tissue repair.

In previous studies, LIPUS has been accepted to promote and improve bone fractures. In addition, the effect of LIPUS on soft tissue repair has attracted much attention, and many studies have been conducted to evaluate the potential effects of LIPUS on hard and soft tissue engineering (25). Chen et al. investigated the effects of LIPUS as supplemental therapy on osteonecrosis of the alveolar bone graft and showed that LIPUS helps to prevent osteonecrosis as a biophysical technique (26). In addition, Toy et al. examined the effects of low pulsed ultrasound waves on bone formation after maxillary expansion in mice. They concluded that cell activation in the LIPUS group was greater than that in the control group, so LIPUS could be accepted as an effective strategy to improve the formation of sutural bone (27).

Since alveolar graft is one of the basic treatments for patients with cleft lip and palate, and due to the importance of the impact of therapeutic methods and techniques on the success of treatments of these patients, in this study we investigated the effect of the usage of low intensity ultrasound on the success of alveolar bone graft.

2- METHODS

2-1. Sampling

Using purpose-oriented sampling and random allocation, in two treatment centers (Orthodontic Department of Mashhad Dental School and Cleft Lip and Palate Clinic of Akbar Children's Hospital), 14 patients with unilateral or bilateral non-syndromic alveolar cleft between the ages of 9 to 13 years, who were candidates for alveolar graft, were selected. The type of intervention was explained to the patients and their parents; and informed consent was obtained from them.

2-2. Patient preparation before surgery

For all patients, standard records including facial and intraoral photographs, panoramic radiographs and study models were prepared. In all patients expansion was considered before surgery. Expansion was performed via quad helix or hyrax. Quad helix was made if the width of the palate was narrow to accommodate hyrax. Hyrax or quad helix were cemented with glass ionomer cement. Patients were instructed to open the hyrax screw every other day and quad helix was made of 0.8 mm SS wire, which was activated 3 mm on each side. The treatment continued until the dental relationships were overcorrected and the appliances were kept in the mouth for another three months for retention.

2-3. Surgery

In both groups, standard alveolar bone grafting was performed, with the mucoperiosteal flap being excised in the cleft region. The nasal floor and buccal flaps were prepared (**Fig. 1**) and the defect area was prepared for transplantation. The autogenous bone (cortical and spongy) was then removed from the iliac region and inserted into the affected area. It should be noted that in patients with bilateral clefts, only one side underwent surgery and the same side was examined. All surgeries were performed by one surgeon.



Fig. 1: Alveolar bone grafting

2-4. Ultrasound application

In seven patients, one week after surgery, the low-intensity pulsed ultrasound waves were used every other day for five minutes at 1 MHz frequency and 100 mW in rotational movements on the skin of the lips of the operated side (**Fig. 2**). This application continued for a total of fifteen sessions in five weeks. The waves were applied with a standard head size of 5 cm² using COMBINED 200 (EME, Italy). This step was performed according to the method proposed by Robertson et al (28).

2-5. Radiographic evaluation

Radiographic examination was performed to evaluate the quality and quantity of bone formed in the graft area. CBCT diagnostic radiography was used and the results were interpreted by an expert oral radiologist.

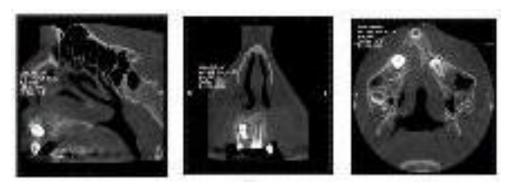


Fig. 2: LIPUS application

CBCT images were taken about one week and three months after surgery. In radiographic examination, bone quantity was measured with two components of height and bone thickness of the area and quality was measured by means of the Hounsfield Unit (HU). All CBCT images were acquired by PLANMECA Promax 3D Max (Helsinki, Finland) with a voxel size of 200 μ m and FOV (field of view) dimension of 100 * 90 * 90 mm³. The PLANMECA Romexis 4.4.3 software was used to examine CBCT images and superimposition (**Fig. 3** and **4**).



(a) (b) (c) **Fig. 3:** CBCT images immediately after alveolar bone graft a: Axial view, b: Frontal view, c: Occlusal view



(a) (b) (c) **Fig. 4:** CBCT images 3 months after alveolar bone graft a: Axial view, b: Frontal view, c: Occlusal view

The superimposition method was that the two CBCT images were matched in three coronal, axial, and sagittal views, and the joint locations including skull base, orbit and other joint points were automatically matched by the software and it was then reviewed by the user for possible troubleshooting. Sites that may have changed during treatment or over time were not considered during superimposition. This method compared the image one week after surgery with the image three months later. The graft in the CBCT was distinct from the surrounding bone.

Mid-sagittal cut was done on the superimposition and images of one week and three months after surgery were displayed separately. On the sides of this mid-sagittal cut, the cuts were struck down 2 mm apart. Height of the graft was measured in different cuts and averaged. Buccolingual thickness was also evaluated and averaged. The mean density was measured in a circle that was adjustable in size. If resorption was seen in the circumferential margins of the circle bone, the smaller circle would be adjusted. However, the diameter of the circle was quite similar in the radiographs after one week and three months after surgery.

2-6. Data analysis

Sample size: According to Alonso et al. (29), at the first type error of 5% and the second type error of 20%, the number of samples in each group was calculated as six patients. However, to increase confidence, the number of samples was increased to seven patients. Statistical methods: Paired t test, Mann-Whitney and Wilcoxon tests were used for statistical analyses via SPSS software version 16.

3- RESULTS

In this study, 14 orthodontic patients with cleft lip and palate, including eight males (57.1%) and six females (42.9%) aged 10 to 13 years were evaluated. First,

demographic information their was reviewed. Seven patients were placed in the intervention group and seven patients in the control group. In the intervention and control groups, three females (42.9%) and four males (57.1%) were present. According to the statistical test results, the gender distribution of the two groups was quite similar (p=1.00). The age range was 10-12 years in the intervention group and 10-13 years in the control group, and there was no significant difference between the groups (p=0.390).

 Table 1 presents the mean, standard
 deviation. minimum, maximum, and median of all the variables in the intervention and control groups, respectively. According to the table, immediately after surgery, the means of sagittal thickness, sagittal height, axial height, and density in the control group were higher than those in the intervention group, but the mean of the axial thickness in the intervention group was higher than that in the control group. Only the difference of density was significant between the two groups (p=0.045). Thus, the two groups were homogeneous for all variables except density.

Table 2presentsthemean,standard deviation. minimum. maximum. and median of all the variables in the intervention and control groups. respectively. As seen, three months later, the means of all variables of sagittal thickness, sagittal height, axial thickness, axial height, and density were higher in the control group than those in the intervention group. However, the difference between the two groups was significant only for sagittal height (p=0.05, 0.021) and was not significant for the other variables.

Table 3 shows that the mean changes of sagittal thickness, sagittal height, and axial thickness in the control group were lower than those in the intervention group, but the mean changes of axial height and density in the intervention group were

lower than those in the control group, but the difference between the two groups was not significant for any of the variables. The two groups were similar in terms of changes during time (one week and three months after surgery).

4-DISCUSSION

Bone repair is one of the most amazing homeostatic activities in the body

(30). Following fracture or transplantation, inflammation, repair, and reconstruction are performed consecutively to restore bone mineralization. The process is slow; so it can cause problems such as fractures and disruption of the treatment process (31). This has prompted researchers to look for different therapies to accelerate the bone healing process (32).

Table-1: Comparing the variables of sagittal thickness, sagittal height, axial thickness, axial	
height and density immediately after surgery between the intervention and control groups	

variable	group	N	mean	Standard deviation	Min	Max	Median	Independent T. test results
Sagittal thickness	intervention	7	9.88	2.28	7.73	14.70	9.53	Z*=0.96
(mm)	control	7	12.40	4.33	6.40	18.37	13.35	P=0.338
Sagittal height	intervention	7	9.84	3.21	6.80	15.23	8.13	T=1.61
(mm)	control	7	12.86	3.80	9.75	20.53	11.20	P=0.134
Axial thickness	intervention	7	10.71	2.40	6.70	13.50	11.18	T=1.01
(mm)	control	7	9.27	2.92	5.00	12.83	10.37	P=0.334
Axial height	intervention	7	8.35	1.83	6.63	12.23	8.05	Z*=1.34
(mm)	control	7	10.64	3.51	5.15	15.15	11.90	P=0.180
Density	intervention	7	266.53	57.62	163.83	349.86	270.45	T=2.24
(Hounsfield unit)	control	7	403.47	151.26	170.71	668.08	390.50	P=0.045

*: Mann-whitney test

Table-2: Comparing sagittal thickness, sagittal height, axial thickness, axial thickness and density three months after the surgery between the intervention and control groups

variable	group	N	mean	Standard deviation	Min	Max	Median	Independent T. test results
Sagittal thickness	intervention	7	6.00	3.67	0.00	10.85	7.20	Z*=0.57 P=0.565
(mm)	control	7	8.65	4.68	5.20	18.27	6.75	
Sagittal height	intervention	7	5.21	2.29	0.00	10.63	4.30	Z*=2.31
(mm)	control	7	9.66	3.55	6.60	15.03	7.80	P=0.021
Axial thickness	intervention	7	5.93	2.84	0.00	8.13	7.10	Z*=0.32 P=0.749
(mm)	control	7	6.40	2.36	4.30	10.45	4.83	
Axial height	intervention	7	5.76	1.72	3.07	8.13	5.63	T=1.30 P=0.219
(mm)	control	7	7.11	2.15	3.80	9.50	7.20	
Density	intervention	7	229.49	120.99	78.79	445.26	204.12	T=1.20
(Hounsfield unit)	control	7	317.67	152.56	155.32	615.37	336.34	P=0.254

*: Mann whitney – test

variable	group	N	mean	Standard deviation	Min	Max	Median	Independent T. test results
Sagittal thickness	intervention	7	3.88	3.03	0.45	9.53	3.85	T=0.07
changes(mm)	control	7	3.75	3.60	0.10	9.46	2.08	P=0.944
Sagittal height	intervention	7	4.62	3.50	1.23	12.07	3.83	Z*=0.70
changes(mm)	control	7	3.20	1.76	0.25	5.50	3.50	P=0.482
Axial thickness	intervention	7	4.78	2.81	1.60	10.25	5.17	T=1.23
changes(mm)	control	7	2.87	3.01	0.35	8.00	1.80	P=0.242
Axial height	intervention	7	2.54	1.37	0.35	4.10	3.23	T=0.96
changes(mm)	control	7	3.53	2.35	0.15	6.30	3.90	P=0.357
Density changes	intervention	7	37.08	76.96	-95.40	106.92	67.52	T=0.79
(Hounsfield unit)	control	7	85.80	143.18	-165.63	262.16	65.19	P=0.443

Table-3: Comparing the changes in the variables of sagittal thickness, sagittal height, axial thickness, axial height and density immediately and three months after surgery between the intervention and control groups

*: Mann-whitney test

One of the procedures that have recently received attention is the use of ultrasound waves. Although many studies have been conducted to evaluate the potential effects of low intensity pulsed ultrasound on soft and hard tissue engineering (25), there has alveolar been no study on cleft reconstructions. To evaluate and compare the therapeutic outcomes in different techniques, evaluation of the amount of bone formed and the height and location of the bone in the alveolar cleft area is necessary. Given the alveolar cleft and considering that there is not enough clinical studies done to evaluate and compare the aforementioned methods, we aimed to investigate the use of lowintensity ultrasound compared to autogenous bone in the repair of alveolar cleft defect. In the present study, similar to other studies, 1 MHz frequency of ultrasound waves was used. The depth of of LIPUS penetration waves is approximately 10 mm, so the application of the head on the skin is appropriate for reaching the bone and graft site (28).

According to the present study, the mean difference between pre- and postoperative variables of sagittal thickness, sagittal height, and axial thickness was less in the control group than that in the intervention group; but the difference between the two groups was not significant for any of the variables. This result indicates that in general, the two groups were similar in terms of changes between the pre- and postoperative methods.

There was no similar research to compare in this area, but our results were not consistent with those studies that showed a positive effect of ultrasound on the fracture healing process (33-35). On the other hand, our results are consistent with the findings of some other researchers reporting no effect for ultrasound on bone repair (36-39).

Differences in the results of research studies investigating the effect of ultrasound on bone tissue and bone fracture healing may be due to differences in the use of different doses and times, the use of ultrasound in human or animal models, and sometimes the use of ultrasound in vitro.

The use of ultrasound in the culture of cells isolated from the body and from other tissues and the general circulation process and various factors including hormones as well as immune and growth factors that flow through the bloodstream and tissue cells can be very different from the effect of ultrasound on the bone tissue in the living organism. Therefore, the positive or negative effects of one or more factors, including ultrasound on the cell or living tissue, can be confirmed in the studies launched in vivo.

Using different methods that produce bone defects or fractures and the use of different intensities of ultrasound cause the results of the research to be different. In selecting the effective intensity of ultrasound, the results have not been uniformly reported and each researcher has suggested a dose according to his/her experience.

Research on the repair of femoral fractures using ultrasound has shown that osteopontin mRNA levels are significantly increased (40). On the other hand, histological examination of tissues after receiving ultrasound at the time of tissue repair and bone healing have shown that the ultrasound effect may be restricted to soft tissues and has no effect on cells located in hard and calcified tissues (41).

5- CONCLUSION

Given the limitations of this study, the following results were obtained:

a) Mean changes in sagittal thickness, sagittal height, and axial thickness were lower in the control group than those in the intervention group, but the difference between the two groups was not significant for any of the variables.

b) In the intervention and control group, mean sagittal thickness, sagittal height, axial thickness, and axial thickness were significantly decreased three months after surgery, but the mean decrease in density was not significant.

c) Low intensity pulsed ultrasound with the frequency of 1 MHz and intensity of 0.1 W/Cm^2 has no significant effect on the clinical success criteria in repairing alveolar defects in patients with cleft palate.

6- REFERENCES

1. Dixon MJ, Marazita ML, Beaty TH, Murray JC. Cleft lip and palate: synthesizing genetic and environmental influences. Nature reviews Genetics. 2011; 12(3):167.

2. Bishara SE. Textbook of orthodontics. 2001.

3. Proffit WR, Fields Jr HW, Sarver DM. Contemporary orthodontics: Elsevier Health Sciences; 2014.

4. Santiago PE, Grayson BH, Cutting CB, Gianoutsos MP, Brecht LE, Kwon SM. Reduced need for alveolar bone grafting by presurgical orthopedics and primary gingivoperiosteoplasty. The Cleft palatecraniofacial journal. 1998; 35(1):77-80.

5. Robertson NR, Jolleys A. An 11-year follow-up of the effects of early bone grafting in infants born with complete clefts of the lip and palate. British journal of plastic surgery. 1983; 36(4):438-43.

6. Steinberg B, Padwa BL, Boyne P, Kaban L. State of the art in oral and maxillofacial surgery: treatment of maxillary hypoplasia and anterior palatal and alveolar clefts. The Cleft palatecraniofacial journal. 1999; 36(4):283-91.

7. Bajaj AK, Wongworawat AA, Punjabi
A. Management of alveolar clefts. Journal of Craniofacial Surgery. 2003; 14(6):8406.

8. Boyne PJ, Sands NR. Combined orthodontic-surgical management of residual palato-alveolar cleft defects. American Journal of Orthodontics. 1976; 70(1):20-37.

9. Boyne PJ. Secondary bone grafting of residual alveolar and palatal clefts. J Oral Surgery. 1972; 30:87-92.

10. Daskalogiannakis J, Ross RB. Effect of alveolar bone grafting in the mixed dentition on maxillary growth in complete unilateral cleft lip and palate patients. The Cleft palate-craniofacial journal. 1997; 34(5):455-8.

11. Dempf R, Teltzrow T, Kramer F-J, Hausamen J-E. Alveolar bone grafting in patients with complete clefts: a comparative study between secondary and tertiary bone grafting. The Cleft palatecraniofacial journal. 2002; 39(1):18-25.

12. Maxson BB, Baxter SD, Vig KW, Fonseca RJ. Allogeneic bone for secondary alveolar cleft osteoplasty. Journal of Oral and Maxillofacial Surgery. 1990; 48(9):933-41.

13. Ma'amon AR, Telfah H. Secondary alveolar bone grafting: the dilemma of donor site selection and morbidity. British Journal of Oral and Maxillofacial Surgery. 2008; 46(8):665-70.

14. Denny AD, Talisman R, Bonawitz SC. Secondary alveolar bone grafting using milled cranial bone graft: a retrospective study of a consecutive series of 100 patients. The Cleft palate-craniofacial journal. 1999; 36(2):144-53.

15. LaRossa D, Buchman S, Rothkopf DM, Mayro R, Randall P. A comparison of iliac and cranial bone in secondary grafting of alveolar clefts. Plastic and reconstructive surgery. 1995; 96(4):789-97.

16. Freihofer HPM, Borstlap WA, Kuijpers-Jagtman AM, Voorsmit RA, van Damme PA, Heidbüchel KL, Borstlap-Engels VM. Timing and transplant materials for closure of alveolar clefts: a clinical comparison of 296 cases. Journal of Cranio-Maxillofacial Surgery. 1993; 21(4):143-8.

17. Booij A, Raghoebar G, Jansma J, Kalk W, Vissink A. Morbidity of chin bone transplants used for reconstructing alveolar defects in cleft patients. The Cleft palate-craniofacial journal. 2005; 42(5):533-8.

18. Walker TW, Modayil PC, Cascarini L, Williams L, Duncan SM, Ward-Booth P.

Retrospective review of donor site complications after harvest of cancellous bone from the anteromedial tibia. British Journal of Oral and Maxillofacial Surgery. 2009; 47(1):20-2.

19. Chen YC, Chen CH, Chen PL, Huang I, Shen YS, Chen CM. Donor site morbidity after harvesting of proximal tibia bone. Head & neck. 2006; 28(6):496-500.

20. Nadal E, Sabás M, Dogliotti P, Espósito R. Secondary alveolar bone grafting: our experience with olecranon bone graft. Journal of Craniofacial Surgery. 2010; 21(2):371-4.

21. Bandow K, Nishikawa Y, Ohnishi T, Kakimoto K, Soejima K, Iwabuchi S, Kuroe K, Matsuguchi T. Low-intensity pulsed ultrasound (LIPUS) induces RANKL, MCP-1, and MIP-1 β expression in osteoblasts through the angiotensin II type 1 receptor. Journal of cellular physiology. 2007; 211(2):392-8.

22. Ikeda T, Yoshizawa S, Tosaki M, Allen JS, Takagi S, Ohta N, Kitamura T, Matsumoto Y. Cloud cavitation control for lithotripsy using high intensity focused ultrasound. Ultrasound in medicine & biology. 2006; 32(9):1383-97.

23. James SL, Ali K, Pocock C, Robertson C, Walter J, Bell J, Connell D. Ultrasound guided dry needling and autologous blood injection for patellar tendinosis. British journal of sports medicine. 2007; 41(8):518-21.

24. Watson T. Ultrasound in contemporary physiotherapy practice. Ultrasonics. 2008; 48 (4):321-9.

25. Tanaka E, Kuroda S, Horiuchi S, Tabata A, El-Bialy T. Low-intensity pulsed ultrasound in dentofacial tissue engineering. Annals of biomedical engineering. 2015; 43(4):871-86.

26. Chen Q, Shi B, Zheng Q. Low intensity pulsed ultrasound therapy: A

potential adjuvant treatment for osteonecrosis of alveolar bone grafting. Journal of research in medical sciences: the official journal of Isfahan University of Medical Sciences. 2013; 18 (3):270-1.

27. Toy E, Ozturk F, Altindis S, Kozacioglu S, Toy H. Effects of lowintensity pulsed ultrasound on bone formation after the expansion of the interpremaxillary suture in rats: a histologic and immunohistochemical study. Australian orthodontic journal. 2014; 30(2):176.

28. Val Robertson, Alex Ward, John Low, Ann Reed. Electrotherapy Explained: Principles and Practice. 4th ed. London, United Kingdom: Elsevier Health Sciences; 2006.

29. Alonso N, Tanikawa DYS, Freitas RdS, Canan Jr L, Ozawa TO, Rocha DL. Evaluation of maxillary alveolar reconstruction using a resorbable collagen sponge with recombinant human bone morphogenetic protein-2 in cleft lip and palate patients. Tissue Engineering Part C: Methods. 2010; 16 (5):1183-9.

30. Mandracchia VJ, Nelson SC, Barp EA. Current concepts of bone healing. Clinics in podiatric medicine and surgery. 2001; 18(1):55-77.

31. Yamada K. Biological effects of low power laser irradiation on clonal osteoblastic cells (MC3T3-E1). Nihon Seikeigeka Gakkai Zasshi. 1991; 65(9):787-99.

32. Trelles M, Mayayo E. Bone fracture consolidates faster with low-power laser. Lasers in surgery and medicine. 1987; 7(1):36-45.

33. Whitecloud T, Cook S, Salkeld S, Ryaby J, editors. Acceleration of spine fusions with a low intensity pulsed ultrasound device. Transactions of the Annual Meeting-Orthopaedic Research Society; 1998: Orthopaedic Research Society. 34. Tsunoda M. Treatment of non-union and delayed-union by low intensity pulsed ultrasound. Clinical calcium. 2003; 13(10):1293-6.

35. Lu H, Qin L, Fok P, Cheung W, Lee K, Guo X, Wong W, Leung K. Low-intensity pulsed ultrasound accelerates bone-tendon junction healing: a partial patellectomy model in rabbits. The American journal of sports medicine. 2006; 34(8):1287-96.

36. Emami A, Petrén-Mallmin M, Larsson S. No effect of low-intensity ultrasound on healing time of intramedullary fixed tibial fractures. Journal of orthopedic trauma. 1999; 13(4):252-7.

37. Spadaro JA, Albanese SA. Application of low-intensity ultrasound to growing bone in rats. Ultrasound in medicine & biology. 1998; 24(4):567-73.

38. Handolin L, Kiljunen V, Arnala I, Kiuru MJ, Pajarinen J, Partio EK, Rokkanen P. No long-term effects of ultrasound therapy on bioabsorbable screw-fixed lateral malleolar fracture. Scandinavian journal of surgery. 2005; 94(3):239-42.

39. Handolin L, Kiljunen V, Arnala I, Kiuru MJ, Pajarinen J, Partio EK, Rokkanen P. Effect of ultrasound therapy on bone healing of lateral malleolar fractures of the ankle joint fixed with bioabsorbable screws. Journal of Orthopaedic Science. 2005; 10(4):391.

40. Ryaby J, Bachner E, Bendo J, Dalton P, Tannenbaum S, Pilla A. Low intensity pulsed ultrasound increases calcium GP Gebauer et al. Journal of Orthopaedic Research. 2002:587-92.

41. Claes L, Willie B. The enhancement of bone regeneration by ultrasound. Progress in biophysics and molecular biology. 2007; 93(1-3):384-98.